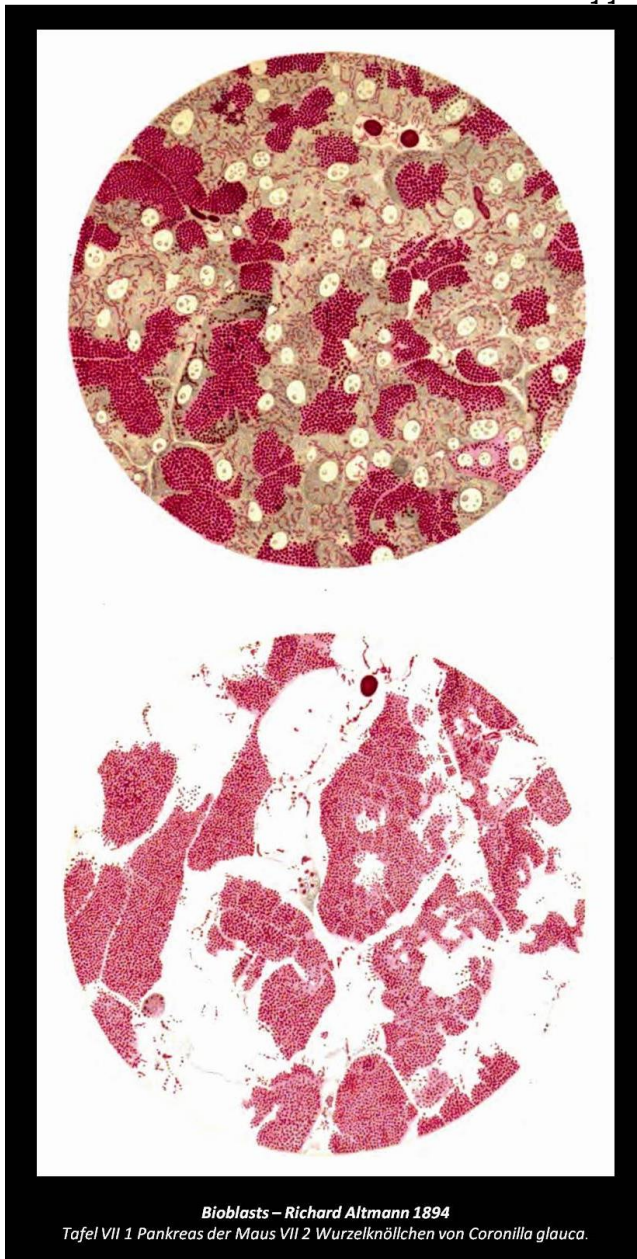


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3
4 **Mitochondrial physiology**
5 **1. Mitochondria and bioblasts**

6
7 MitoEAGLE Task Group*

8
9 *Living Communication:* from Gnaiger *et al* (2020) Bioenerg Commun 2020.1
10



Overview

Richard Altmann (1894):

The protoplasm is a colony of bioblasts. Microorganisms and granula are at an equivalent level and represent elementary organisms, which are found wherever living forces are acting, thus we want to describe them by the common term bioblasts. In the bioblast, that morphological unit of living matter appears to be found.

Mitochondria are oxygen-consuming electrochemical generators that evolved from the endosymbiotic alphaproteobacteria which became integrated into a host cell related to Asgard Archaea (Margulis 1970; Lane 2005; Roger *et al* 2017). Richard Altmann (1894) described the 'bioblasts', which include not only the mitochondria as presently defined, but also symbiotic and free-living bacteria. The word 'mitochondria' (Greek mitos: thread; chondros: granule) was introduced by Carl Benda (1898). Mitochondrion is singular and mitochondria is plural. Abbreviation: mt, as generally used in mtDNA.

Given the multiple roles of mitochondria, it is perhaps not surprising that mitochondrial dysfunction is associated with a wide variety of genetic and degenerative diseases (Falk 2020). Robust mitochondrial function is supported by physical exercise and caloric balance, and is central for sustained metabolic health throughout life. Therefore, a more consistent set of definitions for mitochondrial physiology will increase our understanding of the etiology of disease and

49 improve the diagnostic repertoire of mitochondrial medicine with a focus on protective medicine,
50 evolution, lifestyle, environment, and healthy aging.

51
52 **Updates:**

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54

Abstract56
5758 *Keywords:*59
6061 **Reference: Mitochondrial physiology. 2. Respiratory states and rates**62
63**Mitochondrial structure-function relationships**64 *'For the physiologist, mitochondria afforded the first opportunity for an experimental approach*
65 *to structure-function relationships, in particular those involved in active transport, vectorial*
66 *metabolism, and metabolic control mechanisms on a subcellular level' (Ernster and Schatz 1981).*
67

68

69 Contrary to current textbook dogma, which describes mitochondria as individual organelles,
70 mitochondria form dynamic networks within eukaryotic cells. Mitochondrial movement is
71 supported by microtubules. Mitochondrial size and number can change in response to energy
72 requirements of the cell via processes known as fusion and fission; these interactions allow
73 mitochondria to communicate within a network (Chan 2006). Mitochondria can even traverse cell
74 boundaries in a process known as horizontal mitochondrial transfer (Torralba *et al* 2016).
75 Another defining morphological characteristic of mitochondria is the double membrane. The
76 mitochondrial inner membrane (mtIM) forms dynamic tubular to disk-shaped cristae that
77 separate the mitochondrial matrix, *i.e.*, the negatively charged internal mitochondrial
78 compartment, from the intermembrane space; the latter being enclosed by the mitochondrial
79 outer membrane (mtOM) and positively charged with respect to the matrix.

80 Intracellular stress factors may cause shrinking or swelling of the mitochondrial matrix that
81 can ultimately result in permeability transition (mtPT; Lemasters *et al* 1998). The mtIM contains
82 the non-bilayer phospholipid cardiolipin, which is also involved in the mtOM (Gebert *et al* 2009)
83 but is not present in any other eukaryotic cellular membrane. Cardiolipin has many regulatory
84 functions (Oemer *et al* 2018); it promotes and stabilizes the formation of supercomplexes
85 ($SC_{I_n III_n IV_n}$) based on dynamic interactions between specific respiratory complexes (McKenzie *et al*
86 2006; Greggio *et al* 2017; Lenaz *et al* 2017), and it supports proton transfer on the mtIM from
87 the electron transfer system to F_1F_0 -ATPase (ATP synthase; Yoshinaga *et al* 2016). The mtIM is
88 plastic and exerts an influence on the functional properties of incorporated proteins (Waczulikova
89 *et al* 2007).

90 Mitochondria constitute the structural and functional elementary components of cell
91 respiration. Mitochondrial respiration is the reduction of molecular oxygen by electron transfer
92 coupled to electrochemical proton translocation across the mtIM. In the process of OXPHOS, the
93 catabolic reaction of oxygen consumption is electrochemically coupled to the transformation of
94 energy in the phosphorylation of ADP to adenosine triphosphate (ATP; Mitchell 1961, 2011).
95 Mitochondria are the powerhouses of the cell that contain the machinery of the OXPHOS-
96 pathways, including transmembrane respiratory complexes (proton pumps with FMN, Fe-S and
97 cytochrome *b*, *c*, aa_3 redox systems); alternative dehydrogenases and oxidases; the coenzyme
98 ubiquinone (Q); F_1F_0 -ATPase or ATP synthase; the enzymes of the tricarboxylic acid cycle (TCA),
99 fatty acid and amino acid oxidation; transporters of ions, metabolites and co-factors; iron/sulphur
100 cluster synthesis; and mitochondrial kinases related to catabolic pathways. TCA cycle
101 intermediates are vital precursors for macromolecule biosynthesis (Diebold *et al* 2019). The
102 mitochondrial proteome comprises over 1,200 proteins (Calvo *et al* 2015; 2017), mostly encoded
103 by nuclear DNA (nDNA), with a variety of functions, many of which are relatively well known, *e.g.*,
104 proteins regulating mitochondrial biogenesis or apoptosis, while others are still under
105 investigation, or need to be identified, *e.g.*, mtPT pore and alanine transporter. The mammalian
106 mitochondrial proteome can be used to discover and characterize the genetic basis of
107 mitochondrial diseases (Williams *et al* 2016; Palmfeldt and Bross 2017).
108

109 **Mitochondrial crosstalk**

110

111 Numerous cellular processes are orchestrated by a constant crosstalk between mitochondria and
112 other cellular components. For example, the crosstalk between mitochondria and the endoplasmic
113 reticulum is involved in the regulation of calcium homeostasis, cell division, autophagy,
114 differentiation, and anti-viral signaling (Murley and Nunnari 2016). Mitochondria contribute to
115 the formation of peroxisomes, which are hybrids of mitochondrial and ER-derived precursors
116 (Sugiura *et al* 2017). Cellular mitochondrial homeostasis (mitostasis) is maintained through
117 regulation at transcriptional, post-translational and epigenetic levels (Ling and Rönn 2018;
118 Lisowski *et al* 2018), resulting in dynamic regulation of mitochondrial turnover by biogenesis of
119 new mitochondria and removal of damaged mitochondria by fusion, fission and mitophagy (Singh
120 *et al* 2018). Cell signalling modules contribute to homeostatic regulation throughout the cell cycle
121 or even cell death by activating proteostatic modules, *e.g.*, the ubiquitin-proteasome and
122 autophagy-lysosome/vacuole pathways; specific proteases like LON, and genome stability
123 modules in response to varying energy demands and stress cues (Quiros *et al* 2016). In addition,
124 several post-translational modifications, including acetylation and nitrosylation, are capable of
125 influencing the bioenergetic response, with clinically significant implications for health and
126 disease (Carrico *et al* 2018).

127

128 **The mitochondrial genome**

129

130 Mitochondria of higher eukaryotes typically maintain several copies of their own circular genome
131 known as mitochondrial DNA (mtDNA; hundred to thousands per cell; Cummins 1998), which is
132 maternally inherited in many species. However, biparental mitochondrial inheritance is
133 documented in some exceptional cases in humans (Luo *et al* 2018), is widespread in birds, fish,
134 reptiles and invertebrate groups, and is even the norm in some bivalve taxonomic groups (Breton
135 *et al* 2007; White *et al* 2008). The mitochondrial genome of the angiosperm *Amborella* contains a
136 record of six mitochondrial genome equivalents acquired by horizontal transfer of entire
137 genomes, two from angiosperms, three from algae and one from mosses (Rice *et al* 2016). In
138 unicellular organisms, *i.e.*, protists, the structural organization of mitochondrial genomes is highly
139 variable and includes circular and linear DNA (Zíková *et al* 2016). While some of the free-living
140 flagellates exhibit the largest known gene coding capacity, *e.g.*, jakobid *Andalucia godoyi* mtDNA
141 codes for 106 genes (Burger *et al* 2013), some protist groups, *e.g.*, alveolates, possess
142 mitochondrial genomes with only three protein-coding genes and two rRNAs (Feagin *et al* 2012).
143 The complete loss of mitochondrial genome is observed in the highly reduced mitochondria of
144 *Cryptosporidium* species (Liu *et al* 2016). Reaching the final extreme, the microbial eukaryote,
145 oxymonad *Monocercomonoides*, has no mitochondrion whatsoever and lacks all typical nuclear-
146 encoded mitochondrial proteins, showing that while in 99 % of organisms mitochondria play a
147 vital role, this organelle is not indispensable (Karnkowska *et al* 2016).

148 In vertebrates, but not all invertebrates, mtDNA is compact (16.5 kB in humans) and encodes
149 13 protein subunits of the transmembrane respiratory Complexes CI, CIII, CIV and ATP synthase
150 (F₁F₀-ATPase), 22 tRNAs, and two ribosomal RNAs. Additional gene content has been suggested
151 to include microRNAs, piRNA, smithRNAs, repeat associated RNA, long noncoding RNAs, and even
152 additional proteins or peptides (Rackham *et al* 2011; Duarte *et al* 2014; Lee *et al* 2015; Cobb *et al*
153 2016). The mitochondrial genome requires nuclear-encoded mitochondrially targeted proteins,
154 *e.g.*, TFAM, for its maintenance and expression (Rackham *et al* 2012). The nuclear and the
155 mitochondrial genomes encode peptides of the membrane spanning redox pumps (CI, CIII and
156 CIV) and F₁F₀-ATPase, leading to strong constraints in the coevolution of both genomes (Blier *et al*
157 2001).

158

159 **Mitochondrial respiratory control and regulation**

160

161 The terms metabolic *control* and *regulation* are frequently used synonymously, but are
162 distinguished in metabolic control analysis: “We could understand the regulation as the

163 mechanism that occurs when a system maintains some variable constant over time, in spite of
164 fluctuations in external conditions (homeostasis of the internal state). On the other hand,
165 metabolic control is the power to change the state of the metabolism in response to an external
166 signal" (Fell 1997). Respiratory control may be induced by experimental control signals that exert
167 an influence on: (1) ATP demand and ADP phosphorylation-rate; (2) fuel substrate composition,
168 pathway competition; (3) available amounts of substrates and O₂, *e.g.*, starvation and hypoxia; (4)
169 the protonmotive force, redox states, flux–force relationships, coupling and efficiency; (5) Ca²⁺ and
170 other ions including H⁺; (6) inhibitors, *e.g.*, nitric oxide or intermediary metabolites such as
171 oxaloacetate; (7) signalling pathways and regulatory proteins, *e.g.*, insulin resistance,
172 transcription factor hypoxia inducible factor 1.

173 Mechanisms of respiratory control and regulation include adjustments of: (1) enzyme activities
174 by allosteric mechanisms and phosphorylation; (2) enzyme content, concentrations of cofactors
175 and conserved moieties such as adenylates, nicotinamide adenine dinucleotide [NAD⁺/NADH],
176 coenzyme Q, cytochrome *c*; (3) metabolic channeling by supercomplexes; and (4) mitochondrial
177 density (enzyme concentrations) and morphology (membrane area, cristae folding, fission and
178 fusion). Mitochondria are targeted directly by hormones, *e.g.*, progesterone and glucocorticoids,
179 which affect their energy metabolism (Lee *et al* 2013; Dai *et al* 2013; Gerö and Szabo 2016; Price
180 and Dai 2016; Moreno *et al* 2017; Singh *et al* 2018). Evolutionary or acquired differences in the
181 genetic and epigenetic basis of mitochondrial function (or dysfunction) between individuals; age;
182 biological sex, and hormone concentrations; life style including exercise and nutrition; and
183 environmental issues including thermal, atmospheric, toxic and pharmacological factors, exert an
184 influence on all control mechanisms listed above. For reviews, see Brown 1992; Gnaiger 1993;
185 2001; 2009; 2020; Paradies *et al* 2014; Morrow *et al* 2017.

186 Lack of control by a metabolic pathway, *e.g.*, phosphorylation-pathway, means that there will
187 be no response to a variable activating it, *e.g.*, [ADP]. The reverse, however, is not true as the
188 absence of a response to [ADP] does not exclude the phosphorylation-pathway from having some
189 degree of control. The degree of control of a component of the OXPHOS-pathway on an output
190 variable, such as O₂ flux, will in general be different from the degree of control on other outputs,
191 such as phosphorylation-flux or proton leak flux. Therefore, it is necessary to be specific as to
192 which input and output are under consideration (Fell 1997).

193 Respiratory control refers to the ability of mitochondria to adjust O₂ flux in response to
194 external control signals by engaging various mechanisms of control and regulation. Respiratory
195 control is monitored in a mitochondrial preparation under conditions defined as respiratory
196 states, preferentially under near-physiological conditions of temperature, pH, and medium ionic
197 composition, to generate data of higher biological relevance. When phosphorylation of ADP to ATP
198 is stimulated or depressed, an increase or decrease is observed in electron transfer measured as
199 O₂ flux in respiratory coupling states of intact mitochondria ('controlled states' in the classical
200 terminology of bioenergetics). Alternatively, coupling of electron transfer with phosphorylation
201 is diminished by uncouplers. The corresponding coupling control state is characterized by a high
202 respiratory rate without control by P» (noncoupled or 'uncontrolled state').
203

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347
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350
 351 **Author contributions:** This manuscript developed as an open invitation to scientists and students to join
 352 as coauthors in the bottom-up spirit of COST, based on a first draft written by the corresponding author,
 353 who integrated coauthor contributions in a sequence of Open Access versions. Coauthors contributed to the
 354 scope and quality of the manuscript, may have focused on a particular section, and are listed in alphabetical
 355 order. Coauthors confirm that they have read the final manuscript and agree to implement the
 356 recommendations into future manuscripts, presentations and teaching materials.

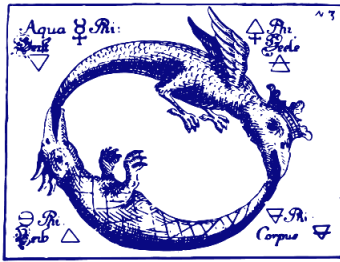


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364
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378

For comments